



TECHNICAL MANUAL

# NADP/NADPH-Glo™ Assay

Instructions for Use of Products  
**G9081 and G9082**

# NADP/NADPH-Glo™ Assay

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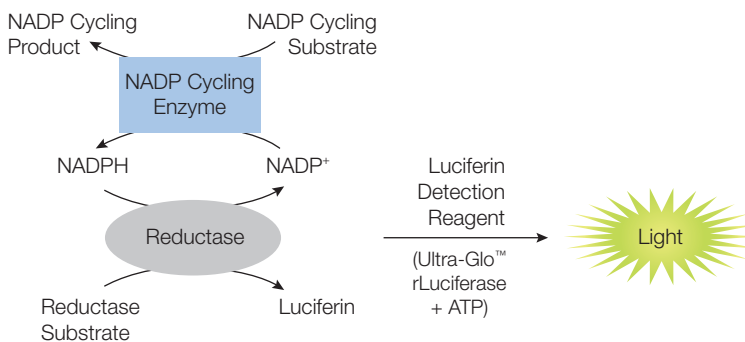
## 1. Description

The NADP/NADPH-Glo™ Assay<sup>(a,b)</sup> is a bioluminescent assay for detecting total oxidized and reduced nicotinamide adenine dinucleotide phosphates (NADP<sup>+</sup> and NADPH, respectively) and determining their ratio in biological samples. NADP<sup>+</sup> and NADPH are critical molecules important for major cellular processes including maintenance of redox balance and biosynthesis of many biomolecules including lipids, nucleotides and amino acids (1,2).

The NADP/NADPH-Glo™ Assay is a homogeneous, single-reagent-addition method to rapidly detect NADP<sup>+</sup> and NADPH in cells and enzymatic reactions and is easily adaptable for inhibitor screening in high-throughput formats.

The NADP Cycling Enzyme is used to convert NADP<sup>+</sup> to NADPH. In the presence of NADPH, the enzyme Reductase reduces a proluciferin reductase substrate to form luciferin. Luciferin is quantified using Ultra-Glo™ Recombinant Luciferase (rLuciferase), and the light signal produced is proportional to the amount of NADP<sup>+</sup> and NADPH in the sample (Figure 1). Cycling between NADP<sup>+</sup> and NADPH by the NADP Cycling Enzyme and Reductase increases assay sensitivity and provides selectivity for the phosphorylated NADP<sup>+</sup> and NADPH compared to the nonphosphorylated forms NAD<sup>+</sup> and NADH.

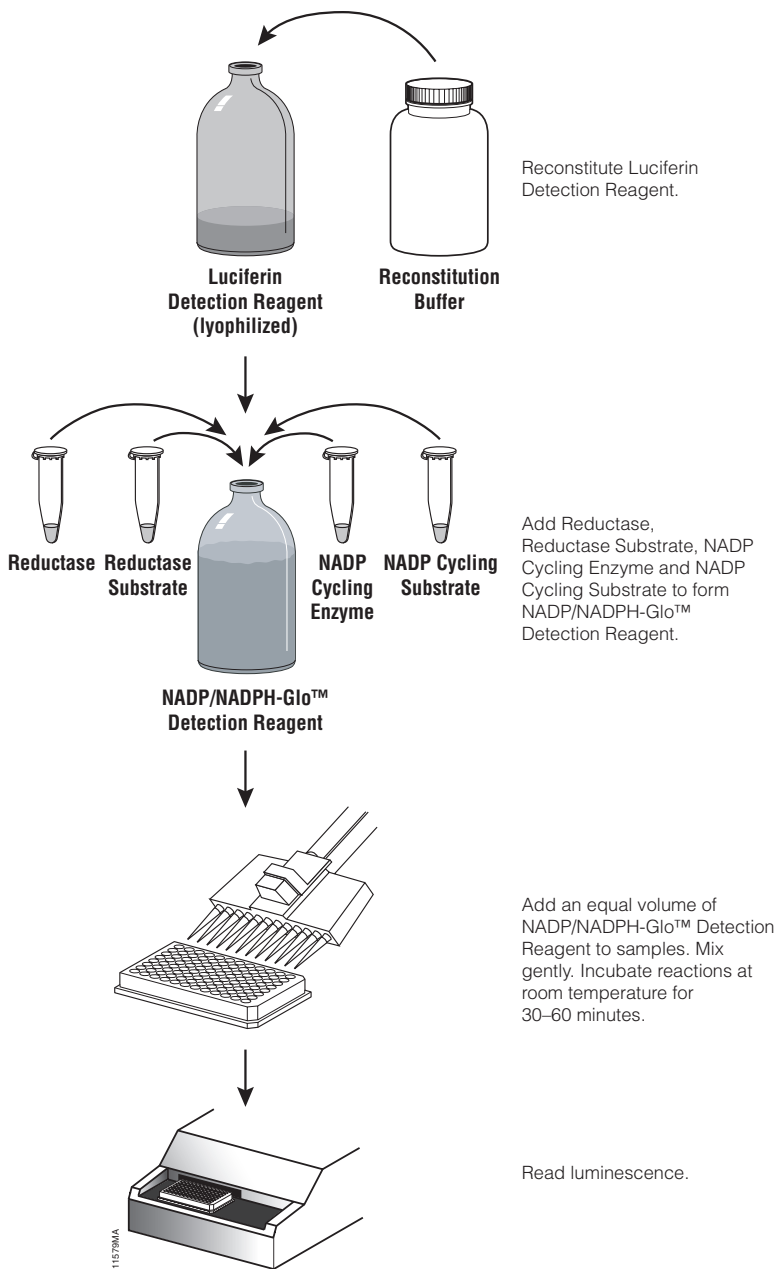
## 1. Description (continued)



**Figure 1. Schematic diagram of the NADP/NADPH-Glo™ Assay technology.** NADP Cycling Enzyme converts NADP<sup>+</sup> to NADPH. In the presence of NADPH, Reductase enzymatically reduces the pro-luciferin Reductase Substrate to luciferin. Luciferin is detected using Ultra-Glo™ rLuciferase, and the amount of light produced is proportional to the amount of NADP<sup>+</sup> and NADPH in a sample.

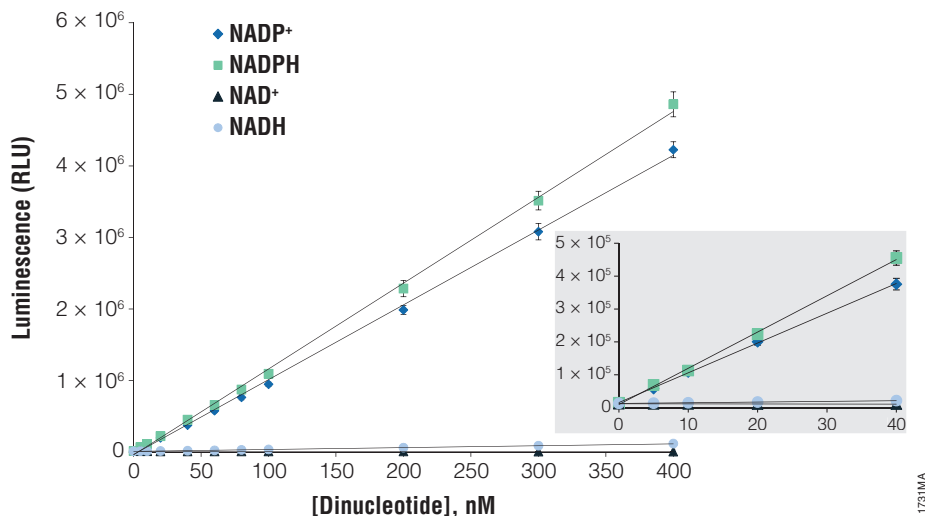
The NADP Cycling Enzyme, Reductase and luciferase reactions are initiated by adding an equal volume of NADP/NADPH-Glo™ Detection Reagent, which contains NADP Cycling Enzyme and Substrate, Reductase, Reductase Substrate and Ultra-Glo™ rLuciferase, to an NADP<sup>+</sup>- or NADPH-containing sample (Figure 2). Detergent present in the reagent lyses cells, allowing detection of total cellular NADP<sup>+</sup> and NADPH in a multiwell format with addition of a single reagent. The one-step protocol is useful for screening changes in total NADP<sup>+</sup> and NADPH levels. An accessory protocol is provided to allow separate measurements of NADP<sup>+</sup> and NADPH and calculation of the NADPH to NADP<sup>+</sup> ratio (Section 5.A). Use the accessory protocol when quantitation of NADP<sup>+</sup> and NADPH is required.

Due to the cycling of the coupled enzymatic reactions, the light signal will continue to increase after adding the NADP/NADPH-Glo™ Detection Reagent to the sample (see Section 6). The luminescent signal remains proportional to the starting amount of NADP<sup>+</sup> and NADPH within the linear range of the assay. The assay has a linear range of 10nM to 400nM NADP<sup>+</sup> and high signal-to-background ratios (Figure 3). The assay is compatible with 96-, 384-, low-volume 384- and 1536-well plates and is well suited to monitor the effects of small molecule compounds on NADP<sup>+</sup> and NADPH levels in enzymatic reactions or directly in cells in high-throughput formats.



**Figure 2. Schematic diagram of the NADP/NADPH-Glo™ Assay protocol.**

1. Description (continued)



**Figure 3. Linear range and specificity of the NADP/NADPH-Glo™ Assay.** Individual purified nicotinamide adenine dinucleotides were assayed following the protocol described in Section 3.C. NADH, NADPH, NAD<sup>+</sup> and NADP<sup>+</sup> stocks were prepared fresh from powder (Sigma Cat. # N6660, N9910, N8285 and N8035, respectively). The concentration of each stock was determined by measuring absorbance at 259nm for NAD<sup>+</sup> and NADP<sup>+</sup> and 340nm for NADH and NADPH using molar extinction coefficients of 18,000 and 6,220, respectively. Stocks were diluted to the indicated concentrations in phosphate-buffered saline (PBS), and 50µl samples at each dinucleotide concentration were incubated with 50µl of NADP/NADPH-Glo™ Detection Reagent in white 96-well luminometer plates. After a 30-minute incubation, luminescence was measured with a GloMax® 96 Microplate Luminometer. Each point represents average luminescence of quadruplicate reactions measured in relative light units (RLU). Error bars are ±1 standard deviation. The limit of detection was approximately 0.5nM for this experiment. The data used to generate this figure are shown in Table 1.

**Table 1. Titration of Purified Dinucleotides.**

Dinucleotide Concentration (nM)	NADP <sup>+</sup>		NADPH	
	Luminescence (RLU)	Signal-to- Background Ratio <sup>1</sup>	Luminescence (RLU)	Signal-to- Background Ratio <sup>1</sup>
400	4,225,190	293.4	4,860,556	328.7
300	3,081,246	213.9	3,514,501	237.7
200	1,986,932	138.0	2,284,997	154.5
100	947,926	65.8	1,095,552	74.1
80	766,097	53.2	874,149	59.1
60	577,148	40.1	659,448	44.6
40	375,652	26.1	455,024	30.8
20	200,187	13.9	223,280	15.1
10	108,373	7.5	112,767	7.6
5	57,926	4.0	68,838	4.7
0	14,403	1.0	14,788	1.0

<sup>1</sup>Signal of sample divided by signal of the 0nM control.

## 2. Product Components and Storage Conditions

PRODUCT	SIZE	CAT. #
<b>NADP/NADPH-Glo™ Assay</b>	<b>10ml</b>	<b>G9081</b>

The system contains sufficient reagents to perform 100 reactions in 96-well plates (100µl of sample + 100µl of NADP/NADPH-Glo™ Detection Reagent), 400 assays in 384-well plates (25µl of sample + 25µl of NADP/NADPH-Glo™ Detection Reagent) or 2,000 assays in low-volume 384-well plates (5µl of sample + 5µl of NADP/NADPH-Glo™ Detection Reagent). Assay volumes can be varied depending on plate format as long as you maintain a 1:1 ratio of sample to NADP/NADPH-Glo™ Detection Reagent. Includes:

- 55µl Reductase
- 55µl Reductase Substrate
- 1 vial NADP Cycling Enzyme (lyophilized)
- 275µl NADP Cycling Substrate
- 1 vial Luciferin Detection Reagent (lyophilized)
- 10ml Reconstitution Buffer

PRODUCT	SIZE	CAT. #
<b>NADP/NADPH-Glo™ Assay</b>	<b>50ml</b>	<b>G9082</b>

The system contains sufficient reagents to perform 500 reactions in 96-well plates (100µl of sample + 100µl of NADP/NADPH-Glo™ Detection Reagent), 2,000 assays in 384-well plates (25µl of sample + 25µl of NADP/NADPH-Glo™ Detection Reagent) or 10,000 assays in low-volume 384-well plates (5µl of sample + 5µl of NADP/NADPH-Glo™ Detection Reagent). Assay volumes can be varied depending on plate format as long as you maintain a 1:1 ratio of sample to NADP/NADPH-Glo™ Detection Reagent. Includes:

- 275µl Reductase
- 275µl Reductase Substrate
- 1 vial NADP Cycling Enzyme (lyophilized)
- 275µl NADP Cycling Substrate
- 1 vial Luciferin Detection Reagent (lyophilized)
- 50ml Reconstitution Buffer

**Storage Conditions:** Store all components below –65°C. Alternatively, store the Reductase Substrate below –65°C and all other components at –30°C to –10°C. Minimize freeze-thaw cycles of all reagents.

### 3. NADP/NADPH-Glo™ Assay Protocol

#### 3.A. Preparing the Luciferin Detection Reagent

1. Thaw the Reconstitution Buffer, and equilibrate the Reconstitution Buffer and lyophilized Luciferin Detection Reagent to room temperature.
2. Transfer the entire contents of the Reconstitution Buffer bottle to the amber bottle of lyophilized Luciferin Detection Reagent.
3. Mix by swirling or inversion to obtain a uniform solution. Do not vortex. The Luciferin Detection Reagent should go into solution easily in less than 1 minute.

**Note:** Store the reconstituted Luciferin Detection Reagent at room temperature while preparing the NADP/NADPH-Glo™ Detection Reagent. If the reconstituted Luciferin Detection Reagent is not used immediately, the reagent can be stored at room temperature (approximately 22°C) for up to 24 hours or dispensed into single-use aliquots and stored at 4°C for up to 1 week or -20°C for up to 3 months with no change in activity.

#### 3.B. Preparing the NADP/NADPH-Glo™ Detection Reagent


Determine the number of NADP/NADPH-Glo™ Assays being performed, and calculate the volume of NADP/NADPH-Glo™ Detection Reagent needed. An equal volume of NADP/NADPH-Glo™ Detection Reagent will be added to each sample containing NADP<sup>+</sup> or NADPH. We recommend preparing extra reagent to compensate for pipetting error. Do not store unused NADP/NADPH-Glo™ Detection Reagent

1. Equilibrate the reconstituted Luciferin Detection Reagent to room temperature.
2. Thaw the Reductase, Reductase Substrate and NADP Cycling Substrate at room temperature or on ice just prior to use. Briefly centrifuge the tubes to bring the contents to the bottom of the tubes, and store on ice.
3. Reconstitute the NADP Cycling Enzyme by adding 275µl of water. Mix by gently swirling the vial, and store on ice.



### 3.B. Preparing the NADP/NADPH-Glo™ Detection Reagent (continued)

4. Prepare the required amount of NADP/NADPH-Glo™ Detection Reagent by adding the volumes of Reductase, Reductase Substrate, NADP Cycling Enzyme and NADP Cycling Substrate indicated in Table 2 per 1ml of reconstituted Luciferin Detection Reagent.

 For best results, we recommend preparing the NADP/NADPH-Glo™ Detection Reagent immediately before use. If necessary, the prepared NADP/NADPH-Glo™ Detection Reagent can be stored at room temperature and used within 6 hours.

**Table 2. Preparing the NADP/NADPH-Glo™ Detection Reagent.**

Component	Volume
Reconstituted Luciferin Detection Reagent	1 ml
Reductase	5µl
Reductase Substrate	5µl
NADP Cycling Enzyme	5µl
NADP Cycling Substrate	5µl

5. Mix by gently inverting five times.
6. Return unused Reductase, NADP Cycling Enzyme and NADP Cycling Substrate to -20°C storage. Return unused Reductase Substrate to storage at less than -65°C. Do not store unused NADP/NADPH-Glo™ Detection Reagent. Minimize the number of freeze-thaw cycles for all reagents.

### 3.C. Protocol

Perform a titration of your particular cell line to determine the linear range and optimal number of cells to use with the NADP/NADPH-Glo™ Assay (see Section 6). Include control wells without cells to determine background luminescence.

This protocol is for a reaction of 50µl of sample and 50µl of NADP/NADPH-Glo™ Detection Reagent in a 96-well plate. The reaction volume can be varied as long as you maintain a 1:1 ratio of sample to NADP/NADPH-Glo™ Detection Reagent. Throughout this manual, sample refers to the starting material such as tissue culture cells.

**Note:** Avoid the presence of DTT and other reducing agents in the samples. Reducing agents will react with the Reductase Substrate and increase background. Also avoid the presence of chelating compounds such as EDTA.

1. Plate cells in a white-walled tissue culture plate and treat with the compounds of interest. The final volume per well should be 50µl.

2. If cells were incubated at 37°C during treatment, remove plate from the incubator, and equilibrate at room temperature for 5 minutes.

**Note:** The assay is compatible with most complete media, making it unnecessary to remove the medium. The medium can be removed and replaced with 50µl of PBS per well if desired.

3. Add 50µl of NADP/NADPH-Glo™ Detection Reagent to each well.
4. Gently and briefly shake the plate to mix and lyse cells.
5. Incubate for 30–60 minutes at room temperature.

**Note:** The light signal will continue to increase with time. Changes in light output can be monitored over time, or luminescence can be measured at a single time point. Be sure to determine the optimal incubation time for your particular application (see Section 6).

6. Record luminescence using a luminometer.

#### 4. General Considerations

##### Plates and Luminometers

Use opaque, white multiwell tissue-culture-treated sterile plates that are compatible with your luminometer (e.g., Corning® 96-well solid white flat-bottom polystyrene TC-treated microplates, Cat.# 3917, or Corning® 384-well low-flange white flat-bottom polystyrene TC-treated microplates, Cat.# 3570). For cultured cell samples, white-walled clear-bottom tissue culture plates (e.g., Corning® 96-well flat clear-bottom white polystyrene TC-treated microplates, Cat.# 3903) are acceptable. If using clear tissue culture plates, you must transfer reactions to white luminometer plates before measuring luminescence. Light signal is diminished in black plates, and increased well-to-well cross talk is observed in clear plates. All standard instruments capable of measuring luminescence are suitable for this assay. Instrument settings depend on the luminometer manufacturer. Use an integration time of 0.25–1 second per well as a guide. Although relative light output will vary with different instruments, variation should not affect assay performance.

##### Temperature

The intensity and stability of the luminescent signal from the NADP/NADPH-Glo™ Assay depend on the reaction rates of the Reductase, NADP Cycling Enzyme and luciferase enzyme. Environmental factors such as temperature affect reaction rates and the intensity of light output. For consistent results, equilibrate the NADP/NADPH-Glo™ Detection Reagent to room temperature (approximately 22°C) before using, and equilibrate assay plates at room temperature for 5 minutes before adding the NADP/NADPH-Glo™ Detection Reagent. Insufficient equilibration may result in a temperature gradient and variability across the plate.

#### **4. General Considerations (continued)**

##### **Chemical Environment**

The chemical environment of the sample containing NADP<sup>+</sup> or NADPH (e.g., cell type, medium and buffer) can affect the Reductase, NADP Cycling Enzyme and luciferase enzymatic rates and light signal intensity. Some media contain ingredients such as pyruvate that can slow down the enzymatic rate. If necessary, increase the incubation time after adding the NADP/NADPH-Glo™ Detection Reagent until sufficient sensitivity is achieved. We recommend testing your particular cell type and medium to determine the optimal cell number and incubation time for your application. The assay is compatible with phenol red.

We recommend a pH of ~7–8 for the NADP<sup>+</sup>- and NADPH-containing samples. Avoid the presence of chelating compounds such as EDTA in the samples. The luciferase reaction requires the divalent magnesium cation, which is included in the Luciferin Detection Reagent. Also, avoid the presence of DTT and other reducing agents in the samples. Reducing agents will react with the Reductase Substrate and increase background.

The NADP/NADPH-Glo™ Assay is compatible with samples containing up to 10% DMSO.

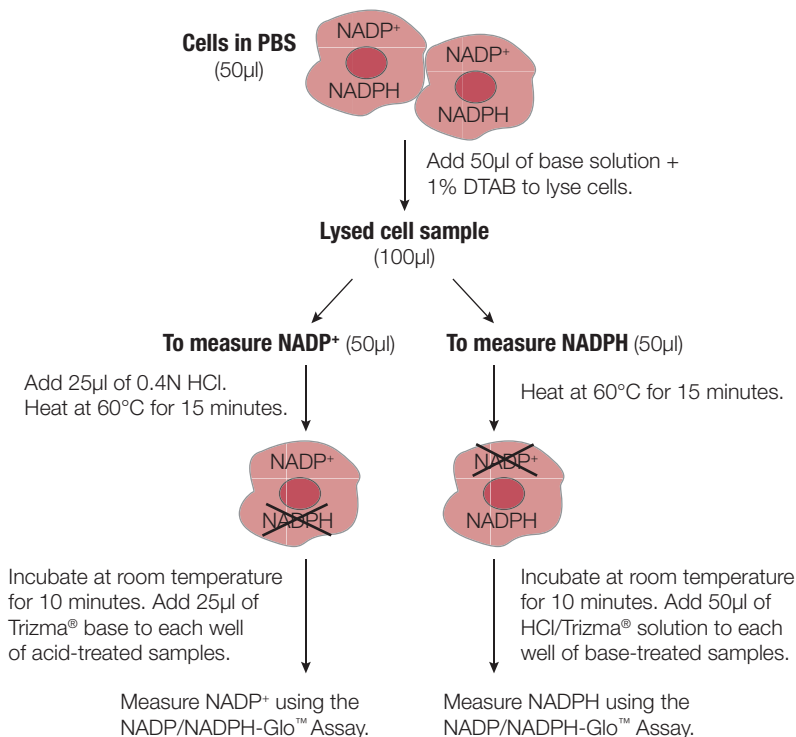
#### **5. Measuring NADP<sup>+</sup> or NADPH Individually**

##### **5.A. Protocol for Sample Preparation**

The protocol to separate oxidized (NADP<sup>+</sup>) and reduced (NADPH) forms takes advantage of the differential stabilities of the forms at acidic and basic pH. In general, oxidized forms are selectively destroyed by heating in basic solution, while reduced forms are not stable in acidic solution (3). Levels of cellular dinucleotides can be individually measured after treatment with acid or base conditions.

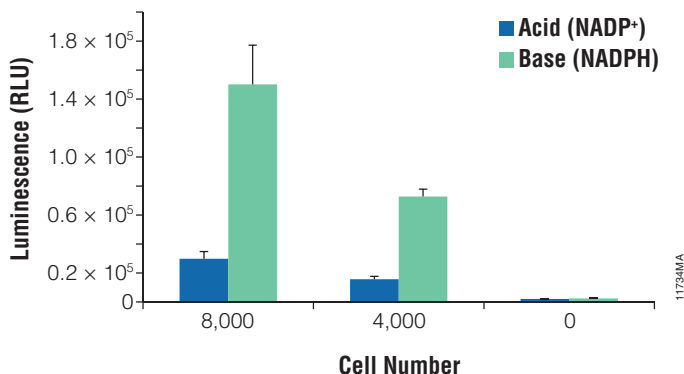
The following sample preparation protocol is recommended for use with the NADP/NADPH-Glo™ Assay to measure NADP<sup>+</sup> and NADPH separately (Figure 4). With this protocol, cells can be processed directly in wells of a 96-well plate. We recommend lysing cells in the preferred base solution with dodecyltrimethylammonium bromide (DTAB), which lyses cells and preserves the stability of the dinucleotides, then splitting the sample into separate wells for acid and base treatments. An advantage of this method is that NADP<sup>+</sup> and NADPH can be measured from one cell sample with in-plate processing. The same treated samples can be used to measure NAD<sup>+</sup> and NADH using the NAD/NADH-Glo™ Assay (Cat. # G9071, G9072).

After sample preparation, all neutralized samples have the same final buffer formulation, which facilitates direct comparison of luminescence values. The direct correlation between luminescence and NADP<sup>+</sup> or NADPH in the samples allows calculation of the NADPH to NADP<sup>+</sup> ratio by dividing luminescence obtained from samples heated in base solution by luminescence obtained from samples heated in acid. Representative data are shown in Figure 5 and Table 3. A standard curve can be generated to quantitate the levels of NADP<sup>+</sup> and NADPH (see Section 5.B).



**Figure 4. Schematic diagram of the sample preparation protocol for measuring NADP<sup>+</sup> and NADPH individually.**

### 5.A. Protocol for Sample Preparation (continued)



**Figure 5. Separate measurement of cellular NADP<sup>+</sup> and NADPH from a single cell sample.** K562 cells were centrifuged and resuspended in PBS at a density of  $1.6 \times 10^6$  cells/ml. After twofold serial dilutions in PBS, 50 $\mu$ l of diluted cells was transferred to each well of a white 96-well plate. Cells were lysed by adding 50 $\mu$ l of bicarbonate base buffer with 1% DTAB and processed as described in Section 5.A. The plate was weighed before and after heating to quantify evaporation. A  $\leq 2\%$  change in weight was observed, indicating minimal sample loss due to evaporation. Twenty microliters of each neutralized sample, containing the indicated number of cell equivalents, was transferred to a 384-well plate, and the NADP/NADPH-Glo™ Assay protocol was performed as described in Section 3.C. The average of quadruplicate reactions is plotted. Error bars are  $\pm 1$  standard deviation.

**Table 3. Calculation of the NADPH to NADP<sup>+</sup> Ratio.**

	Cell Number <sup>1</sup>		
	8,000	4,000	0
Luminescence of acid-treated samples (NADP <sup>+</sup> ) (RLU)	29,790	15,698	2146
Luminescence of base-treated samples (NADPH) (RLU)	150,100	72,727	2511
Ratio of NADPH to NADP <sup>+</sup>	5.0	4.6	

<sup>1</sup>Number of cell equivalents in 20 $\mu$ l of neutralized sample combined with 20 $\mu$ l of NADP/NADPH-Glo™ Detection Reagent.

## Materials to Be Supplied by the User

(Solution compositions are provided in Section 8.)

- phosphate-buffered saline (e.g., Sigma Cat.# D8537 or Gibco Cat.# 14190)
- base solution: bicarbonate base buffer or 0.2N NaOH  
**Note:** Two base solutions, the bicarbonate base buffer and 0.2N NaOH, have been tested in the protocol and perform similarly. For 100 samples, 4.8ml of either base solution is required.
- 0.4N HCl
- base solution with 1% DTAB (bicarbonate base buffer with 1% DTAB or 0.2N NaOH with 1% DTAB)
- 0.5M Trizma® base
- HCl/Trizma® solution

This protocol is for assaying cells in 50µl of PBS per well in 96-well white luminometer plates. Each well of cells is split into two samples: One sample is treated with acid to quantify NADP<sup>+</sup>, and the other is treated with base to quantify NADPH (see Figure 4). When plating cells, reserve half of the wells on the plate for splitting samples. Alternatively, use a second plate when splitting samples.

1. Prepare the Luciferin Detection Reagent as described in Section 3.A.
2. To each well of cells in 50µl of PBS, add 50µl of base solution with 1% DTAB.
3. Briefly mix plate on a plate shaker to ensure homogeneity and cell lysis.
4. Remove 50µl of each sample to an empty well for acid treatment. To these samples, add 25µl of 0.4N HCl per well; these wells contain the acid-treated samples. The original sample wells are the base-treated samples; do not add 0.4N HCl to those wells.
5. Cover the plate, and incubate all samples for 15 minutes at 60°C.
6. Equilibrate the plate for 10 minutes at room temperature.
7. Add 25µl of 0.5M Trizma® base to each well of acid-treated cells to neutralize the acid.
8. Add 50µl of HCl/Trizma® solution to each well containing base-treated samples.  
**Note:** At this point, the total volume per well is 100µl. To perform the NADP/NADPH-Glo™ assay, add 100µl of NADP/NADPH-Glo™ Detection Reagent directly to each well in Step 10. Alternatively, remove a portion of the sample to another plate before adding an equal volume of NADP/NADPH-Glo™ Detection Reagent (e.g., transfer 20µl of sample to a 384-well plate, and add 20µl of NADP/NADPH-Glo™ Detection Reagent).
9. Prepare the NADP/NADPH-Glo™ Detection Reagent as described in Section 3.B.

### 5.A. Protocol for Sample Preparation (continued)

10. Add an equal volume of NADP/NADPH-Glo™ Detection Reagent (e.g., 100µl) to each well.
11. Gently shake the plate to mix.
12. Incubate for 30–60 minutes at room temperature.
13. Record luminescence using a luminometer.

**Note:** The oxidized form (NADP<sup>+</sup>) is selectively destroyed by heating in basic solution, while the reduced form (NADPH) is not stable in acidic solution. Thus, luminescence from acid-treated samples is proportional to the amount of NADP<sup>+</sup>. Luminescence from base-treated samples is proportional to the amount of NADPH.

### 5.B. Generating a Standard Curve

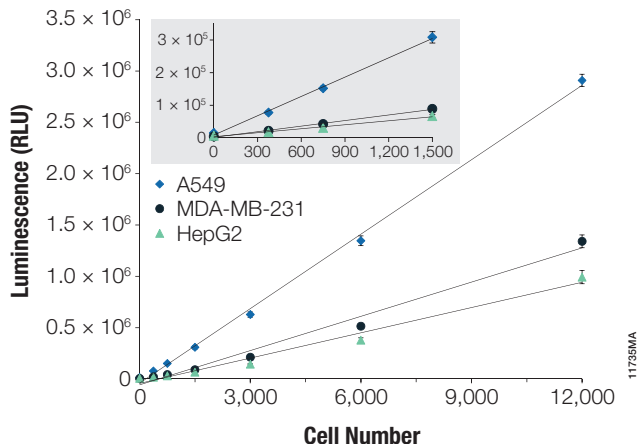
A standard curve allows conversion of luminescence (in RLU) to NADP<sup>+</sup> or NADPH concentration by directly comparing luminescence from samples to the light signals generated from purified NADP<sup>+</sup> or NADPH. For the standard curve, we recommend using purified NADP<sup>+</sup> to prepare a concentrated stock of 2mM NADP<sup>+</sup> in PBS. (If Sigma Cat.# N8035 is used, the stock solution can be prepared directly in the vial.) Immediately before the assay, prepare standard samples at the desired concentrations by diluting the 2mM NADP<sup>+</sup> stock in the same buffer used to prepare the experimental samples, as pH and some buffer components can affect the light signal (see Section 4). If experimental samples were generated using the sample preparation protocol in Section 5.A, dilute the NADP<sup>+</sup> in a mixture of equal volumes of PBS, base solution with 1% DTAB, 0.4N HCl and 0.5M Trizma® base. Assay each standard sample on the same plate as the experimental samples. Include control wells that lack NADP<sup>+</sup>.

For each point on the standard curve, calculate average luminescence, and subtract average luminescence of the blank reactions (reactions at 0nM NADP<sup>+</sup>) to obtain net luminescence. Use the net luminescence values to generate the standard curve and perform linear regression analysis. Interpolate the amount of NADP<sup>+</sup> or NADPH by comparing net luminescence values of the experimental samples to the values in the standard curve.

## 6. Establishing the Linear Range with Cells

Luminescence is directly proportional to cell number over the linear range of the NADP/NADPH-Glo™ Assay. The NADP/NADPH-Glo™ Assay is compatible with many cell types and media. However, absolute light signal intensity and linear range depend on specific cell type and medium (Figure 6).

We recommend testing your particular cell type and medium to determine the linear range, optimal cell number and optimal incubation time for your application.



**Figure 6. Linear relationship between light signal and cell density.** The indicated number of cells were assayed in medium supplemented with 10% fetal bovine serum (FBS) (F-12K for A549 cells, DMEM for MDA-MB-231 cells and EMEM for HepG2 cells) in wells of 96-well white plates. Fifty microliters of NADP/NADPH-Glo™ Detection Reagent was added to 50µl of each cell type at each dilution. After a 60-minute incubation, the light signal was measured in a GloMax® 96 Microplate Luminometer. The values represent the average of quadruplicate reactions, and error bars are ±1 standard deviation. The CV values were ≤5%.

Due to the cycling of the coupled enzymatic reactions, the light signal will continue to increase after adding the NADP/NADPH-Glo™ Detection Reagent to the sample. Changes in light output can be monitored over time, or luminescence can be measured at a single time point. Optimal light signal will usually be generated within 30–60 minutes. The linear range changes with time, and at later time points, samples at higher cell numbers may be out of the linear range of the assay. Light output remains proportional to the amount of NADP<sup>+</sup> or NADPH in the sample until all of the Reductase Substrate is converted to luciferin.

**Note:** If a stable light signal is preferred (for example, when batch processing multiwell plates), the increase in signal after adding the NADP/NADPH-Glo™ Detection Reagent can be stopped at any time by adding the reductase inhibitor menadione. Add 10% of the reaction volume (i.e., 10µl to a 100µl reaction) of 2.75mM menadione prepared in 20% DMSO for a final concentration of 0.25mM menadione.



## 7. References

1. Pollak, N., Dölle, C. and Ziegler, M. (2007) The power to reduce: Pyridine nucleotides—small molecules with a multitude of functions. *Biochem. J.* **402**, 205–18.
2. Icard, P. and Lincet, H. (2012) A global view of the biochemical pathways involved in the regulation of the metabolism of cancer cells. *Biochim. Biophys. Acta* **1826**, 423–33.
3. Lowry, O.H., Passonneau, J.V. and Rock, M.K. (1961) The stability of pyridine nucleotides. *J. Biol. Chem.* **236**, 2756–9.

## 8. Composition of Buffers and Solutions

### Base solution with 1% DTAB

To one of the base solutions (i.e., bicarbonate base buffer or 0.2N NaOH), add 20% DTAB to a final concentration of 1% (v/v). For example, to 4.75ml of base solution, add 0.25ml of 20% DTAB.

### Bicarbonate base buffer

100mM	sodium carbonate
20mM	sodium bicarbonate
10mM	nicotinamide
0.05%	Triton® X-100

The pH of the prepared buffer will be approximately 10–11.

### 20% DTAB

Prepare a 20% DTAB (Sigma Cat.# D8638) solution in water. Warm the solution in a 37°C water bath to completely solubilize the DTAB. Store at room temperature or –20°C.

### 0.4N HCl

Prepare 0.4N HCl from a concentrated stock solution such as 1N HCl by diluting with water. No pH adjustment is required.

### HCl/Trizma® solution

Add equal volumes of 0.4N HCl and 0.5M Trizma® base. Mix by vortexing.

### 0.2N NaOH

Prepare 0.2N NaOH from a concentrated stock solution such as 1N NaOH by diluting with water to 0.2N. No pH adjustment is required.

### 0.5M Trizma® base

Dissolve 12.1g Trizma® base powder (Sigma Cat.# T1503) in 200ml of water. The final pH will be approximately 10.7. No pH adjustment is required.

## 9. Related Products

### Energy Metabolism Assays

<b>Product</b>	<b>Size*</b>	<b>Cat #</b>
NAD(P)H-Glo™ Assay	10ml	G9061
NAD/NADH-Glo™ Assay	10ml	G9071
Glucose Uptake-Glo™ Assay	5ml	J1341
Lactate-Glo™ Assay	5ml	J5021
Malate-Glo™ Assay	5ml	JE9100
Pyruvate-Glo™ Assay	5ml	J4051
Glucose-Glo™ Assay	5ml	J6021
Glycogen-Glo™ Assay	5ml	J5051
Glutamine/Glutamate-Glo™ Assay	5ml	J8021
Glycerol-Glo™ Assay	5ml	J3150
Triglyceride-Glo™ Assay	5ml	J3160
Cholesterol/Cholesterol Ester-Glo™ Assay	5ml	J3190
BCAA-Glo™ Assay	5ml	JE9300
BHB-Glo™ Assay	5ml	JE9400
Metabolite-Glo™ Detection System	5ml	J9030
ROS-Glo™ H2O2 Assay	10ml	G8820
GSH/GSSG-Glo™ Assay	10ml	V6611

\*Additional sizes available.

### Cell Viability Assays

<b>Product</b>	<b>Size*</b>	<b>Cat #</b>
CellTiter-Glo® 2.0 Cell Viability Assay	10ml	G9241
CellTiter-Glo® 3D Cell Viability Assay	10ml	G9681
RealTime-Glo™ MT Cell Viability Assay	100 assays	G9711
CellTiter-Fluor™ Cell Viability Assay	10ml	G6080

\*Additional sizes available.



## 9. Related Products (continued)

### Cytotoxicity and Apoptosis Assays

Product	Size*	Cat #
LDH-Glo™ Cytotoxicity Assay	10ml	J2380
CellTox™ Green Cytotoxicity Assay	10ml	G8741
Caspase-Glo® 3/7 Assay System	2.5ml	G8090
RealTime-Glo™ Annexin V Apoptosis and Necrosis Assay	100 assays	JA1011

\*Additional sizes available.

## 10. Summary of Changes

The following changes were made to the 1/24 revision of this document:

1. Updated patent statements.
2. Changed font and cover image.
3. Made minor text edits.

<sup>(a)</sup>Patent Pending.

<sup>(b)</sup>U.S. Pat. Nos. 9,273,343 and 9,951,372, European Pat. No. 2751089, Japanese Pat. No. 6067019 and other patents pending.

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